

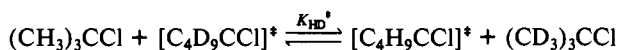
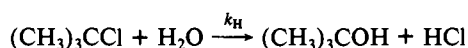
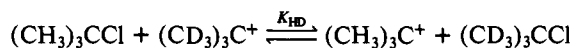
An Equilibrium Deuterium Isotope Effect of Surprising Magnitude

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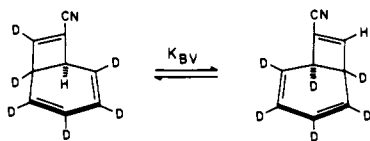
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Any equilibrium isotope effect measures the difference between local force fields at two different sites. The heavier isotope always prefers the more constrained site. Deuterium equilibrium isotope effects can also be used to calibrate the more frequently measured secondary deuterium kinetic isotope effects. For example, K_{HD} for *tert*-butyl- d_9 cation exchange (2.36, 25 °C) is hardly distinguishable from k_H/k_D for hydrolysis² (2.39), i.e., from K_{HD} .³ It follows that the structure of the hydrolytic transition state— $[C_4H_9CCl]^*$ —closely resembles that of a *tert*-butyl cation.⁴ The



differences between these two different kinds of isotope effects become larger and more variable in [3,3]-sigmatropic shifts; so also do the corresponding differences in transition-state structures.⁵

In the absence of such thermodynamic calibration, secondary deuterium kinetic isotope effects are occasionally difficult to distinguish from small primary isotope effects. For example, a solvolytic β -secondary deuterium isotope effect greater than ca. 1.2 (per deuterium atom) is occasionally taken to suggest the incursion of rate-determining deprotonation^{6a} or hydrogen participation.^{6b} Nevertheless, larger equilibrium isotope effects than this have been reported. In the cyanobicyclo[4.2.0]octatrienes- d_6 , for example, deuterium prefers a bridgehead (B) to a vinyl (V) site by $K_{BV} = 1.55$ (6).⁷



We now report the preference of deuterium for an α -chloro site (α) to a bridgehead site (B) of $K_{\alpha B} = 1.48$ (4) and of this same α -chloro site to a vinyl site (V) of $K_{\alpha V} = 3.0$ (1). To the best of our knowledge, this latter isotope effect exceeds any previously reported value, at least for deuterium equilibration between two different carbon sites.⁸

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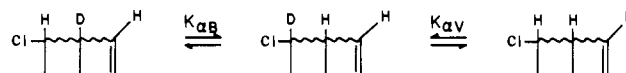
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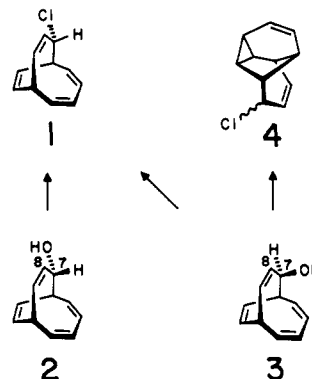
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(7) (a) Paquette, L. A.; Kitching, W.; Heyd, W. E.; Meisinger, R. H. *J. Am. Chem. Soc.* **1974**, *96*, 7371-7372. (b) The authors explicitly recognize this to be a maximum value, the product of K_{BV} with a secondary kinetic deuterium isotope effect of unknown magnitude. The latter can be estimated as ca. 1.08, whence $K_{BV} \approx 1.44$.



Both isotope effects were obtained from samples of *anti*-7-chlorobicyclo[4.3.2]undecatetraene- d_1 ($1-d_1$). The vinyl site (V) is here an average over eight nonequivalent locations, and the bridgehead site (B) is an average over two. As we report elsewhere,⁹ **1** is obtained as the exclusive product of anti alcohol (**2**) and as coproduct of syn alcohol (**3**), the second coproduct being an epimeric mixture of arminenyl chlorides (**4**). When these latter chlorides (**4**) were obtained from specifically deuterated **3**, their ¹H NMR spectrum revealed a deuterium distribution that was essentially random. The corresponding distribution in $1-d_1$, however, was subtly different.



Entries 3-12 of Table I show that the vinyl proton areas of $1-d_1$ are consistently greater than anticipated for a completely random distribution. The α -proton and bridgehead proton areas are correspondingly lower, albeit to different extents. These discrepancies persist despite differences in the stereochemistry of the alcohol precursor in the original locations of the deuterium label, in the reaction conditions, or in the purification of the product. They cannot be due to partial dedeuteration by extraneous protons, because concordant results were obtained in the presence of both HCl (oxalyl chloride conditions) and excess triethylamine (mesyl chloride conditions). They vanish when a ¹³C label is used⁹ or in the absence of any label at all (entries 1 and 2). These discrepancies must therefore be measuring a deuterium isotope effect.

Is this also an equilibrium deuterium isotope effect? It need not be if chloride ion capture of equilibrating carbocations were irreversible. The small secondary kinetic isotope effect, to be expected of such capture, might then be amplified by incomplete carbocation equilibration.¹⁰ Even larger effects might then also be "induced" by deprotonating side reactions.¹¹ Fortunately, no such reactions could be detected in the ¹H NMR spectra, even of "crude" product. To guarantee the reversibility of chloride ion capture is more difficult, at least in the absence of specifically deuterated $1-d_1$.

That equilibration does extend to the covalent chlorides was first suggested by related experiments with specifically deuterated arminenyl chlorides ($4-d_1$). These equilibrate rapidly by a carbocation mechanism, even in chloroform solution at ambient temperatures.¹² Because the bicyclic chloride (**1**) hydrolyzes more rapidly than do the arminenyl chlorides (**4**),⁹ one expects the isotopic equilibration of **1** also to be more rapid. Confirmatory evidence was provided by entry 13 of Table I. The otherwise slow

(8) The autoprotolysis constant of H₂O exceeds that of D₂O by a factor of 6.88. See: Kingerly, R. W.; LaMer, V. K. *J. Am. Chem. Soc.* **1941**, *63*, 3256-3261.

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Table I. Fractional ¹H NMR Areas and Equilibrium Constants (35 °C) of *anti*-7-Chlorobicyclo[4.3.2]undecatetraene-*d*₁ (*1-d*₁)

| structural assignment: δ _H , ppm ^a areas calcd for random distribution: | | vinyl 6.10-5.45 | α 4.80-4.70 | bridgehead 3.65-3.40 | | | |
|---|---|------------------------------------|----------------|-------------------------|--------------------|------------------------------|------------------------------|
| | | 0.727 | 0.091 | 0.182 | Isolated | | |
| precursor | reaction conditions | fractional areas obsd ^b | | | yield ^c | K _{αB} ^b | K _{αV} ^b |
| 1. <i>syn</i> -3- <i>d</i> ₀ | SOCl ₂ , pyr/ether | 0.725 (3) | 0.092 (2) | 0.184 (1) | | | |
| 2. <i>anti</i> -2- <i>d</i> ₀ | SOCl ₂ , pyr/ether | 0.730 (3) | 0.091 (1) | 0.179 (2) | | | |
| 3. <i>syn</i> -3-8- <i>d</i> ₁ | SOCl ₂ , pyr/ether | 0.749 (6) | 0.083 (4) | 0.168 (4) | 98% | 1.1 (3) | 2.7 (6) |
| 4. <i>syn</i> -3-8- <i>d</i> ₁ | MesCl Et ₄ NCl, Et ₃ N | 0.748 (3) | 0.079 (2) | 0.173 (2) | 97% | 1.6 (2) | 3.3 (3) |
| 5. <i>syn</i> -3-7- <i>d</i> ₁ | SOCl ₂ , pyr/ether | 0.763 (5) | 0.073 (5) | 0.167 (2) | 100% | 1.4 (2) | 6 (2) |
| 6. <i>anti</i> -2-8- <i>d</i> ₁ | SOCl ₂ , pyr/ether | 0.746 (6) | 0.085 (2) | 0.169 (6) | 98% | 1.00 (6) | 2.2 (2) |
| 7. <i>anti</i> -2-8- <i>d</i> ₁ | MesCl, Et ₄ NCl, Et ₃ N | 0.743 (4) | 0.086 (2) | 0.171 (3) | 94% | 1.0 (3) | 2.0 (4) |
| 8. <i>anti</i> -2-7- <i>d</i> ₁ | SOCl ₂ /pentane | 0.754 (3) | 0.073 (5) | 0.172 (5) | 94% | 1.7 (3) | 3.8 (6) |
| 9. <i>anti</i> -2-7- <i>d</i> ₁ | SOCl ₂ /pyr, ether | 0.757 (7) | 0.076 (2) | 0.167 (2) | 100% | 1.6 (1) | 4.3 (5) |
| 10. <i>anti</i> -2-7- <i>d</i> ₁ | (CICO) ₂ /PhH(vac) ^d | 0.747 (4) | 0.080 (3) | 0.172 (1) | <i>e</i> | 1.8 (3) | 3.6 (7) |
| 11. <i>anti</i> -2-7- <i>d</i> ₁ | (CICO) ₂ /PhH | 0.750 (1) | 0.076 (2) | 0.174 (1) | <i>e</i> | 1.5 (2) | 3.0 (7) |
| 12. <i>anti</i> -2-7- <i>d</i> ₁ | (CICO) ₂ /PhH | 0.744 (3) | 0.079 (3) | 0.176 (2) | <i>f</i> | 1.83 (9) | 2.9 (3) |
| 13. <i>anti</i> -1 (from 12) | SnCl ₄ , CH ₂ Cl ₂ | 0.743 (3) | 0.077 (3) | 0.181 (3) | <i>f</i> | 2.2 (1) | 3.1 (3) |
| | weighted mean ^g | 0.749 (1) | 0.079 (1) | 0.172 (1) | | 1.48 (4) | 3.0 (1) |

^a Continuous wave spectra in CDCl₃ at 90 MHz except where otherwise specified. ^b Mean and standard deviations in the last digit as obtained from 3-5 scans. Except where otherwise specified, all samples were purified via HPLC. ^c Prior to HPLC purification. ^d Vacuum line techniques were used. Inert atmosphere syringe techniques were used elsewhere. ^e Analysis without prior HPLC purification. ^f FT spectra at 80 MHz. ^g $\bar{x} = \Sigma(x_i/s_i)/\Sigma(1/s_i)$ and $s = 1/(\Sigma 1/s_i^2)^{1/2}$.

structural isomerization of *1-d*₁ to *4-d*₁ was catalyzed by stannic chloride, and the process was halted after 60% completion. Recovered unreacted *1-d*₁ should then have had ample opportunity for reversible chloride dissociation to be complete. Because the resulting isotopic distribution remained unchanged, it seems reasonable to identify its persistence with chemical equilibrium.

Equations 1 and 2 were then used to define and evaluate the two equilibrium constants.¹³ Each *n*_{*i*} represents the mole fraction

$$K_{\alpha B} \equiv \frac{2n_{\alpha}}{n_B} = \frac{a_V - 9a_{\alpha} + a_B}{a_V + a_{\alpha} - 4a_B} \quad (1)$$

$$K_{\alpha V} \equiv \frac{8n_{\alpha}}{n_V} = \frac{4a_V - 36a_{\alpha} + 4a_B}{-a_V + 4a_{\alpha} + 4a_B} \quad (2)$$

of one isotopic isomer with its deuterium atom at site *i*; the *a*_{*i*} are the corresponding observed areas. Because the algebraic form of these equations attenuates experimental precision, nonrandom error propagation was avoided by evaluating *K*_{αB} and *K*_{αV} separately for each NMR scan. To the extent that precursors were incompletely deuterated, 1.48 and 3.0 should be regarded as *lower limits* of the true equilibrium constants. One recalls too that *K*_{αV} is an average over 8 nonequivalent sites; some of them might well require larger equilibrium isotope effects than the rest.¹⁴

Why are these isotope effects so large? Molecular models reveal no apparent steric constraints about the α hydrogen, but constraints might well be present nonetheless. Nucleophiles attack the corresponding ketone almost exclusively from the anti direction.¹⁵ Perhaps the preference for bridgehead to vinyl deuteration (*K*_{BV} = *K*_{αV}/*K*_{αB} = 2.0) is greater than that in the cyanobicyclo[4.2.0]octatrienes, because there the CH bond is also a less constrained cyclobutyl CH bond.¹⁶

A more interesting possibility is that these isotope effects only appear to be large for want of adequate comparison. If so, they might profitably encourage discovery of still larger effects in molecules that provide correspondingly greater contrasts in local force fields. Empirical calibration of secondary deuterium kinetic

isotope effects could then rest more securely upon a wider range of structurally well-defined equilibrium isotope effect data.

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Supplementary Material Available: A brief derivation of eq 1 and 2 (1 page). Ordering information is given on any current masthead page.

Formation of *N*-Phenylheme in the Hemolytic Reaction of Phenylhydrazine with Hemoglobin

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The formation of a green pigment in erythrocytes of animals treated with phenylhydrazine was described by Hoppe-Seyler in 1885.¹ Pigment formation has subsequently been found to be intimately associated with the precipitation of hemoglobin in the form of Heinz bodies and with the ensuing hemolytic anemia.² Despite a century of continuous scrutiny, however, the nature of the green chromophore and the mechanism by which it is formed remain unknown. The green substance was actually isolated in

(13) See supplementary material. A more general derivation will be presented elsewhere. The critical reader can verify eq 1 and 2 at the extremes; e.g., *K*_{αB} = *K*_{αV} = 1 when *a*_V = 8, *a*_α = 1, and *a*_B = 2.

(14) 270-MHz NMR spectra resolved the two bridgehead protons and showed them to be of equal area. The vinyl protons remained incompletely resolved, even at 600 MHz.

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